

EFFECTS OF X-RAYS UPON HAPLOID AND DIPLOID EMBRYOS OF HABROBRACON¹

A. M. CLARK AND C. J. MITCHELL²

*Dept. of Biological Sciences, University of Delaware, Newark, Delaware, and
Marine Biological Laboratory, Woods Hole, Mass.*

Correlations between radiosensitivity and ploidy, the number of chromosome sets, have been made by Stadler (1929), Müntzing (1941), Fröier, Gelin and Gustafsson (1941) and Smith (1943, 1946) for cereals; by Latarjet and Ephrussi (1949) for the yeast, *Saccaromyces*; by Whiting and Bostian (1931), Clark and Kelly (1950), and Clark and Mitchell (1951) for *Habrobracon*; and by Lamy and Muller (1939) for *Drosophila*. These studies, with the exception of the last, have been consistent in demonstrating, for the material tested, that there is a greater resistance with higher ploidy and the results have been taken as evidence that x-radiation damage is primarily chromosomal. That there is less chance for homozygous deficiencies to occur in polyploids than in diploids and that all deficiencies in haploids would be lethal, have been given as explanations. Lamy and Muller found that diploid and triploid *Drosophila* x-rayed as embryos are equally radiosensitive and they assume that the deleterious effects in this case are largely non-chromosomal ("physiological").

The senior author and associates have been studying the effects of x-rays on haploids and diploids of the parasitic wasp, *Habrobracon*, during different stages of its life cycle in order to determine to what extent genome number can be correlated with radiosensitivity. Comparison of radiosensitivity during the pupal, prepupal and larval stages has shown that diploids are more resistant than haploids. The present paper reports on radiosensitivity during the early embryonic stages.

MATERIAL AND METHODS

In *Habrobracon*, haploid males arise from unfertilized eggs and diploids (male and female) from fertilized eggs. Whiting (1943) has established the sex-determining mechanism and has shown that cultures can be obtained that contain (1) only haploid males, (2) haploid males and diploid females, (3) haploid males, diploid females and diploid males. In the present study comparison is made between (1) and (2). Stocks No. 33 and No. 17-o¹ (ivory) were used. Cultures of haploid embryos were obtained from No. 33 unmated mothers while cultures of haploid and diploid embryos ("mixed" cultures) were obtained from No. 33 females mated to No. 17-o¹ males. Progeny of mated females normally consist of about 60 per cent diploids and 40 per cent haploids.

Embryos of known ages were x-rayed. They were then (a) placed in syracuse dishes containing a mineral oil ("Nujol") and observed for hatchability or (b)

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placed upon food (paralyzed *Ephestia* larvae) in order that survivors could develop into adults. All cultures were kept at 30° C. At this temperature eggs hatch about 30 hours after being laid. The larvae in mineral oil were counted 40 hours after oviposition and the incidence of hatchability noted. Eggs placed upon *Ephestia* larvae were observed at various stages of development and comparison of sex ratios was made between control and treated cultures from mated females.

All microscopical studies were made upon whole mounts of embryos fixed in Kahle's fluid and stained with the Rafalko modification of the Feulgen technique.

For x-ray experiments a dual-tube self-rectifying outfit with a simultaneous cross firing technique was used. The secondary voltage was 182 kv.; the tube was 25 ma.; the output intensity was 110 r per minute. There was an inherent filter equivalent to 0.2 mm. copper. X-radiation was obtained at the Marine Biological Laboratory, Woods Hole, Massachusetts.

RESULTS

Embryos treated between one and three hours of age

As determined by microscopical study of whole mounts, untreated embryos between one and three hours of age were found not to have progressed beyond cleavage. There is a syncytium at this stage with the nuclei rapidly undergoing synchronous mitosis. For the majority of the eggs the number of nuclei found was over fifty. A range from the pronuclear stage to embryonic stages having over

TABLE I
Hatchability ratios for eggs from mated and unmated females
(Age of embryos when x-rayed, 1-3 hours)

Dose (r)	Haploids (from unmated ♀ ♀)		Haploids	Diploids*	Diploids (estimated) hatchability
			(from mated ♀ ♀)		
	No. eggs	Hatchability	No. eggs	Hatchability	
0	573	.93±.01	1071	.92±.01	.91±.01
27.5	91	.97±.02	131	.93±.02	.90±.04
55	148	.91±.02	263	.87±.02	.75±.03
110	263	.85±.02	433	.47±.02	.22±.03
165	220	.69±.03	168	.31±.03	.07±.02
220	331	.51±.03	319	.18±.02	.00
275	137	.27±.04	146	.05±.02	
330	195	.10±.02	225	.04±.02	
440	142	.03±.01	161	.02±.01	

* Adult female ratio in controls .60 (729/1226 adults).

250 nuclei was observed but these extremes were exceptional. Eggs from mated and unmated females were in the same stages of development.

Hatchability ratios for eggs of this age at time of treatment demonstrate that those from mated females are more sensitive than those from unmated (Table I). This difference is clearly shown after doses from 110 r to 275 r. For example,

after treatment with 110 r, the hatchability for eggs from mated females is $0.47 \pm .02$ as compared with $0.85 \pm .02$ for eggs from unmated mothers. Untreated eggs from mated and unmated females do not differ in hatchability (Table I). Since the incidence of female offspring from mated control mothers was found to be 0.60 on the basis of 1226 adults counted, these data can be used for deriving the hatchability of diploid eggs (Table I).

Since the hatchability data (Table I) indicate that haploids are more radio-resistant, the ratio of adult females $\left(\frac{\text{♀♀}}{\text{Total}}\right)$ from x-rayed eggs of mated mothers should be lower than from control eggs. This is substantiated when comparison is made of the ratios of haploid males and diploid females from x-rayed and control cultures (Table II). At all doses there is a marked decrease in the ratio of females. For example, after treatment with 220 r, 33 males and 6 females were obtained ($0.15 \pm .06$) as compared with 46 males and 60 females in the untreated group ($0.57 \pm .05$).

Reference to Table II will show that there is a decrease in hatchability with increase in dose and that this occurs for eggs from both mated and unmated females.

TABLE II
Hatchability and eclosion ratios for offspring from mated females
(Age of embryos when x-rayed, 1-3 hours)

Dose (r)	No. Eggs	No. adults		Larvae Eggs	Adults Larvae	♀ ♀ Total
		♂ ♂	♀ ♀			
0	146	46	60	.84±.03	.86±.03	.57±.05
55	116	43	43	.81±.04	.91±.03	.50±.05
110	101	26	7	.55±.05	.59±.07	.21±.07
165	284	42	8	.18±.02	.98±.02	.16±.05
220	404	33	6	.11±.02	.86±.05	.15±.06

Once the eggs have hatched, however, there appear to be no further deleterious effects as shown by comparison of ratios of adults, larvae at different doses. There is no developmental lag in survivors and the adults show no structural abnormalities. Death due to irradiation occurs before hatching.

Embryos that were x-rayed (220 r) during cleavage (one-three hours of age) were fixed at an age when somites were normally present (12-15 hours of age) in the control embryos. Examination of whole mounts showed that some of these treated embryos had formed somites and appeared normal. The majority, however, were either in cleavage or early blastema. Nuclei in the cleavage stages were in interphase and were very much enlarged, being up to four times the diameter of untreated nuclei. Many were clumped together forming dark patches within the egg. The large number of nuclei in treated embryos which had died may suggest that cleavage continued after x-radiation. In some eggs, nuclei could not be found, due, perhaps, to disintegration of chromatin material. A sufficient number of eggs from haploid and mixed cultures was not prepared to make a quantitative microscopical comparison between these groups.

TABLE III

*Hatchability and eclosion ratios for offspring from mated females
(Age of embryos when x-rayed, 3-4 hours)*

Dose (r)	No. eggs	No. adults		$\frac{\text{Larvae}}{\text{Eggs}}$	$\frac{\text{Adults}}{\text{Larvae}}$	$\frac{\text{♀ ?}}{\text{Total}}$
		♂ ♂	♀ ♀			
0	197	53	102	.89±.02	.89±.02	.66±.04
220	442	48	81	.34±.02	.86±.03	.63±.04

Embryos treated between three and four hours of age

Embryos of three-four hours of age from mated mothers were treated with 220 r and were allowed to develop into adults. The sex ratios of adults emerging were compared with those from control eggs (Table III). The incidence of females from control and treated eggs is not significantly different. This indicates that haploids and diploids are equally radiosensitive when treated at this stage of development. There are no developmental effects of an injurious nature after hatching as shown by the ratios of adults/larvae. Most of the three-hour old control embryos have completed cleavage and have hundreds of nuclei within the egg. These nuclei are migrating or have migrated to the periphery of the egg (blastema stage).

Embryos treated between 13 and 15 hours of age

Older embryos in the stage when somites are present that are treated with x-rays may not show injury during the egg stages. However, deleterious effects may be seen during later stages of development. Embryos, 13-15 hours of age, were x-rayed with either 722 r or 1444 r. Comparison of the number of larvae obtained showed that there is no decrease in hatchability over the controls (Table IV). The number of adults, however, is markedly decreased for those treated with 1444 r. These groups tended to show a lag in development as larvae and to remain as larvae after the controls had become adults. Some of the larvae did not grow to full size, but continued development to the pupal stage without spinning a cocoon.

TABLE IV

*Hatchability and eclosion ratios for offspring from mated and unmated females
(Age of embryos when x-rayed, 13-15 hours)*

Dose (r)	Mothers	No. eggs	No. larvae	No. adults		No. dying as larvae	$\frac{\text{Larvae}}{\text{Eggs}}$	$\frac{\text{Adults}}{\text{Larvae}}$
				♂ ♂	♀ ♀			
0	mated	42	30	11	12	2	.71	.77
722	mated	80	64	27	20	8	.80	.73
1444	mated	53	40	4	3	25	.75	.17
0	unmated	42	39	35		4	.93	.90
722	unmated	47	43	36		7	.92	.84
1444	unmated	50	44	12		24	.88	.27

There are insufficient data to state whether haploids and diploids are affected differentially when irradiated at 13–15 hours of age.

DISCUSSION

Comparisons of radiosensitivity among individuals differing in ploidy have appeared in the literature in an attempt to evaluate the extent of chromosomal ("genetic") and cytoplasmic ("physiological") injury. Since differences in the number of chromosome sets are the most obvious distinguishing characteristics, the occurrence of a differential lethal effect has been taken as a criterion that nuclear injury has taken place. It is less obvious, but pertinent, that different metabolic conditions may exist in diploids and polyploids. Investigations herein reported of x-ray effects for haploids and diploids of *Habrobracon* suggest that radiosensitivity cannot be correlated with genome number at all stages in the life cycle. The differential radiosensitivity between haploids and diploids depends upon the stage of development at which they are irradiated. Haploids are more resistant than diploids during cleavage, equally resistant immediately following cleavage, and less resistant than diploids during the larval, prepupal, and pupal stages. One might inquire, therefore, if a differential effect between diploids and polyploids is an adequate criterion for distinguishing between chromosomal and cytoplasmic injury.

Most of the investigators who have reported a differential effect upon survival have found that the individuals with the greater number of chromosome sets are more resistant. Some of them (Müntzing, 1941; Latarjet and Ephrussi, 1949; Clark and Kelly, 1950; Clark and Mitchell, 1951) have suggested that (1) chromosome breakage occurs with the resultant loss of acentric fragments, and (2) there is a greater chance for individuals with higher ploidy to retain unbroken chromosomes that could compensate for the homologous fragments that are lost.

There are some observations that are difficult to reconcile with interpretations of radiation injury purely in terms of a chromosome breakage hypothesis. Studies on diploid and polyploid cereals have correlated gross observations on survival with the number of chromosome breaks (Fröier, Gelin and Gustafsson, 1941; Smith, 1943, 1946). These investigators have shown that polyploids, even though more resistant than diploids, have more anaphase bridges and chromosome fragments than comparable diploids. Fröier, Gelin and Gustafsson (1941) have shown that the germination and sprouting ability of polyploids were unimpaired even when 50 per cent of the mitoses had bridges and fragments, but the growth of the diploids was impaired with much less nuclear disturbance. Further, they showed that at high doses (50,000–60,000 r) nuclei of *Avena sativa*, a hexaploid, are still able to divide even if the chromosomes are fragmented to the extent of discontinuity. Marshak and Bradley (1944) have shown that mitotic inhibition is inversely proportional to the number of chromosomes, but independent of chromosome length. They suggest that this indicates an effect upon the centromeres.

If a chromosomal basis for injury is to be postulated, then a different mechanism must be used to explain the fact that haploid embryos of *Habrobracon* are more resistant than diploids when treated during cleavage. The observations by Fröier, Gelin and Gustafsson (1941) and Smith (1943, 1946) on cereals and by Bishop (1950) on *Tradescantia*, that the number of chromosome breaks following x-radiation is proportional to the number of chromosomes, would suggest that the

diploid nuclei of *Habrobracon* incur twice as many chromosome breaks as comparable haploid nuclei. One might reason that the diploids would be more sensitive than the haploids because they have received twice as much injury per nucleus. It has already been noted, however, that polyploids with more chromosome breaks per nucleus are more resistant than diploids with fewer chromosome breaks per nucleus. Differences between the stages of development that allow haploids to be more resistant at one stage and more sensitive at another should perhaps be considered. A number of obvious differences exist between the cleavage stage and the later post-embryonic stages of *Habrobracon*. During cleavage there is a syncytium with the nuclei dividing rapidly and synchronously throughout the embryo while in later embryonic and the post-embryonic stages cells are present with active proliferation being restricted to localized regions. There is little differentiation in the embryos during cleavage. Whether or not these differences have anything to do with the reversal in radiosensitivity is not known.

If differential radiosensitivity is to be taken as a criterion of chromosomal injury, then the equivalent radiosensitivity of haploid and diploid embryos when x-rayed immediately after cleavage must be taken to mean that extra-chromosomal factors are involved. The radioresistance is increasing rapidly at this time and, therefore, a slight lag in development between haploids and diploids could result in a large difference in radiosensitivity. Kelly (unpublished), using 50 per cent hatchability as the criterion for comparative lethality, has shown that haploid embryos increase in resistance from a dose of about 200 r during cleavage to 7000 r in about four hours. Lamy and Muller (1939) found that diploid and triploid *Drosophila* when treated as embryos were equally radiosensitive. They explained this by assuming that death was due to a "non-genetic" ("physiological") type of injury. The data reported in the present paper suggest that the *Drosophila* embryos were x-rayed after cleavage.

Embryos x-rayed during cleavage or in early blastema are arrested in development during the egg state or not at all. Post-embryonic development is normal. Embryos x-rayed at a later stage, however, may show a post-embryonic lag in development and become arrested at the larval stage. Dent and Amy (1950) treated embryos and larvae of *Habrobracon* with P^{32} . They found a lag and an arresting of development at the larval stage. Henshaw and Henshaw (1933) and Packard (1935) have determined the sensitivity of *Drosophila* eggs when x-rayed during different stages of embryonic development using hatchability as the criterion of radiosensitivity. Since in *Habrobracon* older embryos when x-rayed may hatch normally but may show deleterious effects after hatching, it seems that hatchability alone is not an adequate measure of radiosensitivity. Kelly (unpublished) showed that an x-ray dose of 30,000 r had no effect on the hatchability of older embryos of *Habrobracon*. Such embryos, however, would not have developed beyond the larval stage.

Although its significance is not known, the observation that nuclei of embryos arrested during cleavage are considerably larger following irradiation is in agreement with the reports of enlarged nuclei by Mottram (1933) for *Colpidium*, Jensen's rat sarcoma and bean roots, and by Harrington and Koza (1951) for grasshopper embryos.

These data on *Habrobracon* show that radiosensitivity cannot be correlated with genome number throughout the life cycle. It is difficult, therefore, to pose a single

generalization that will explain these diverse data from the standpoint of chromosomal injury. It may also be difficult to explain them on the basis of a quantitative difference of some chemical constituent. It is possible that there is no single "most-radiosensitive" mechanism, but that the relative sensitivities of the cellular materials change during development and that different mechanisms may be primarily involved at different stages.

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SUMMARY

1. Comparison of radiosensitivity of haploids and diploids of *Habrobracon* shows that the stage of development at which the animals are x-rayed is important in determining the relative sensitivity between these groups. When embryos are x-rayed during cleavage, haploids are more resistant than diploids; when embryos are x-rayed immediately after cleavage has been completed (blastema stage), haploids and diploids are equally radiosensitive.

2. Embryos x-rayed during cleavage or early blastema are deleteriously affected during the egg stage or not at all. Those that hatch complete post-embryonic development normally. Older embryos when x-rayed may hatch, but post-embryonic development is slowed down and many of the individuals are arrested in development as larvae. Hatchability, therefore, is not an adequate criterion of radiosensitivity for older embryos.

3. Embryos that are x-rayed during cleavage and fail to hatch are arrested in cleavage or in early blastema. The nuclei are arrested at interphase and become enlarged up to four times the diameter of untreated nuclei.

4. Since the differential radiosensitivity between haploids and diploids depends upon the stage of development at which they are irradiated, it is difficult to pose a single hypothesis that will account for these facts. It seems reasonable to consider that the relative sensitivities of the cellular materials change during development and that different mechanisms may be primarily involved at different stages.

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